

Design, synthesis and biological evaluation of a macrocyclic discodermolide/dictyostatin hybrid†

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A 22-membered macrocyclic discodermolide/dictyostatin hybrid has been designed and synthesised; biological evaluation against a range of human cancer cell lines revealed significant levels of growth inhibition.

Discodermolide (**1**, Fig. 1), originally isolated from the deep-sea sponge *Discodermia dissoluta*, displays potent antiproliferative activity against a wide range of human cancer cell lines and inhibits the growth of drug-resistant solid tumours.¹ It shares a similar microtubule-stabilising mechanism to that of Taxol, while having a greater tubulin binding affinity,² and has progressed into clinical development as a novel anticancer agent.³ Furthermore, the synergistic combination of Taxol and discodermolide induces tumour regressions and suppresses angiogenesis in animal models of ovarian carcinoma, supporting their potential use together in cancer therapeutics.⁴

Similarly, dictyostatin (**2**) displays further elevated levels of growth inhibition across the same wide range of human cancer cell lines, and has emerged as a new microtubule-stabilising agent with

promising anticancer properties.^{5,6} Strong structural similarities exist between discodermolide and dictyostatin, particularly with regard to stereochemical homology, as determined by our recent configurational assignment of the latter structure using detailed NMR analysis,^{5c} suggesting that they interact in a similar fashion with the Taxol binding site on β -tubulin.^{5d,6} Thus, building on initial encouraging findings reported by the Curran group,^{7a} constraining the conformation of the more flexible open-chain structure of discodermolide into the macrocyclic ring motif of dictyostatin might provide active hybrids of these marine-sponge-derived polyketides. Herein, we report the synthesis of the designed 22-membered macrolide **3**, incorporating the full C2–C24 linear sequence of discodermolide and the (*Z*)-enoate of dictyostatin, and that it shows significant growth inhibitory activity against human cancer cell lines.

At the outset, we considered it essential to achieve a suitable overlay of the energetically preferred conformation of our designed hybrid with that of discodermolide. A 10 000 step Monte Carlo conformational search was performed using MacroModel (Version 8.0) with the MM2* force field and a Born/surface area (GB/SA) water solvent model. The calculated global minimum for analogue **3** (see the ESI†) correlates well with the X-ray crystal structure of discodermolide,^{1a} as shown in the overlay in Fig. 1. The match for the C9–C26 region (corresponding to C7–C24 of discodermolide) region is particularly striking. Consequently, if the tubulin-bound conformation of discodermolide resembles its X-ray structure, the constrained macrocyclic analogue **3** may possess a similar binding affinity and cytotoxicity to that of the natural product.

Our synthetic strategy leading to analogue **3** was adapted from previous work on the total synthesis of discodermolide.⁸ A complex aldol coupling of enone **4** and aldehyde **5**, derived from previously prepared advanced intermediates, would thus be used to assemble a suitable linear precursor for macrolactonisation (Scheme 1).

Synthesis of the enone **4** started from bis-PMB ether **6**,^{8a} which incorporates the stereochemistry required for the C11–C26 region (Scheme 2). Selective removal of the primary PMB ether was achieved with $\text{BCl}_3 \cdot \text{DMS}$,⁹ followed by oxidation of the resulting alcohol with TEMPO/PhI(OAc)₂ and Still–Gennari olefination¹⁰ to give enone **4** (73%). Synthesis of the aldehyde partner **5** began with the selective primary oxidation of diol **7**,^{8b,c} again making use of TEMPO/PhI(OAc)₂, followed by a Still–Gennari olefination to provide (*Z*)-enoate **8** (60%). Removal of the PMB ether,¹¹ bis-silylation of the corresponding diol with TBSOTf/2,6-lutidine and DIBALH reduction of the methyl ester generated allylic alcohol **9** (48%). Finally, PMB ether formation with PMBTCA/Sc(OTf)₃, selective primary TBS cleavage and Dess–Martin oxidation

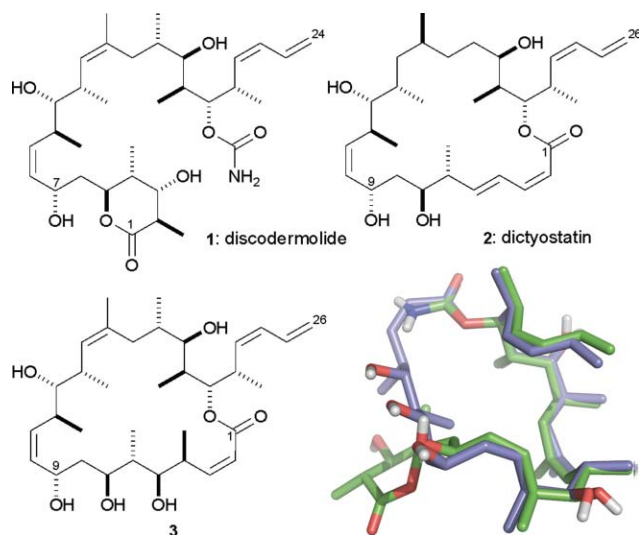
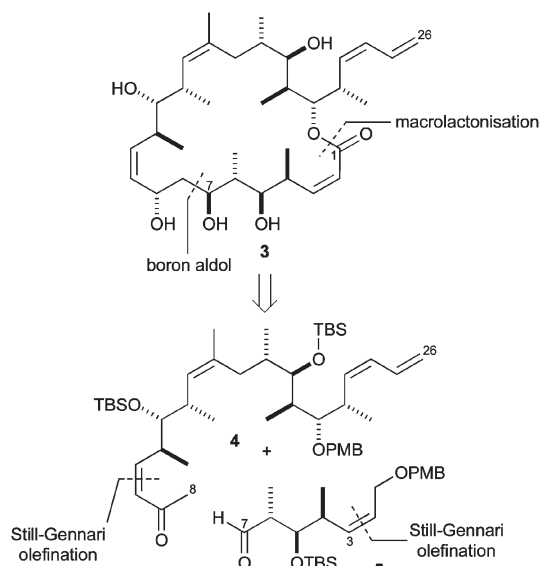


Fig. 1 Structures of discodermolide, dictyostatin and designed hybrid **3**. Overlay of the lowest energy conformation of hybrid **3** (purple) and the X-ray structure of discodermolide (green).

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† Electronic supplementary information (ESI) available: Experimental and modelling details, and ¹H/¹³C NMR data for compounds **3** and **10–15**. See DOI: 10.1039/b615122a

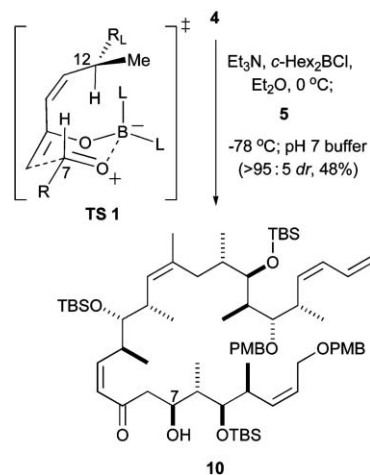


Scheme 1 Retrosynthetic analysis of hybrid analogue 3.

provided aldehyde 5 (68%) in readiness for the pivotal aldol coupling step.

Enolisation of enone 4 with *c*-Hex₂BCl/Et₃N in Et₂O, and subsequent addition of aldehyde 5, gave adduct 10 with >95 : 5 dr in favour of the desired (*7S*)-adduct (48%, Scheme 3). This anti-Felkin–Anh outcome is consistent with our earlier work on 1,6-induction in similar boron aldol reactions, and can be rationalised by invoking the favoured transition state model **TS 1**.^{8b,c}

Reduction of β -hydroxyketone 10 proved problematic, affording mixtures of epimeric alcohols at C9 when subjected to standard Evans–Saksena conditions (Scheme 4).¹² An improvement was found by following the same protocol as previously employed for the reduction of similarly troublesome β -hydroxyketones^{8b} using stoichiometric quantities of (*R*)-CBS and BH₃·THF,¹³ which provided the desired isomer 11 in a 75 : 25 dr.¹⁴ The 1,3-*anti* diol 11 was protected as its acetone; subsequent oxidative PMB ether cleavage (with DDQ) and sequential oxidation of the resultant

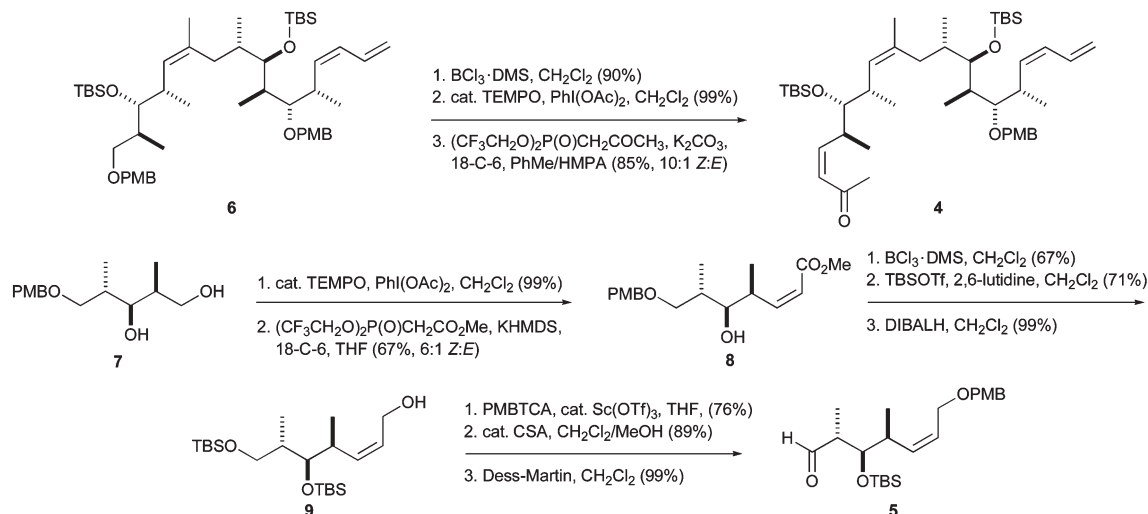


Scheme 3 Key boron aldol coupling.

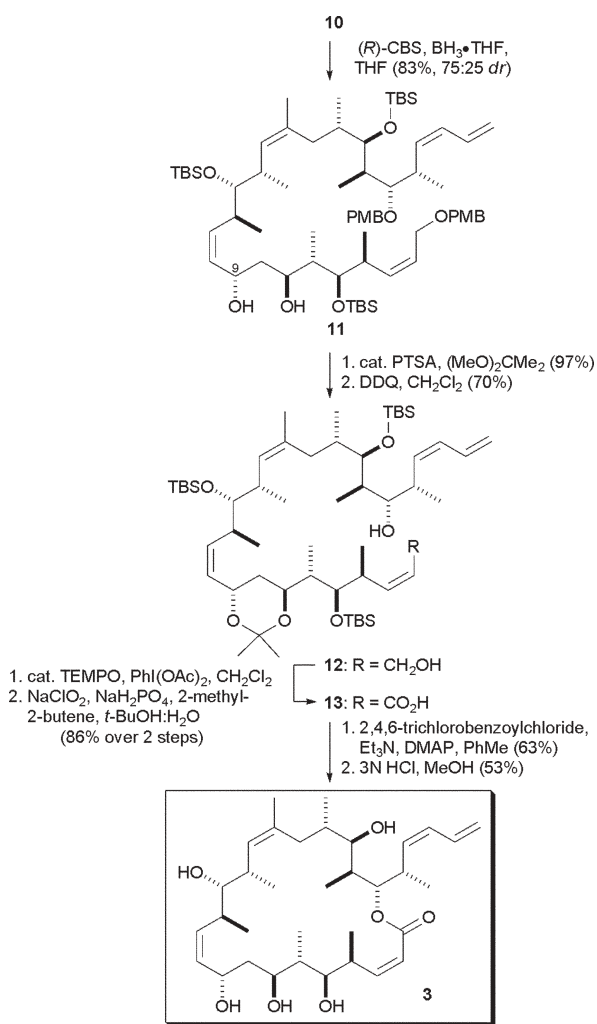
alcohol 12 with TEMPO/PhI(OAc)₂ then NaClO₂/NaH₂PO₄ provided *seco*-acid 13 (58%).

Finally, a Yamaguchi macrolactonisation of 13 (65%) and global deprotection using 3M HCl in MeOH (53%) afforded the 22-membered macrolide 3 after HPLC purification. Full proton and carbon assignments were carried out using COSY and HMQC NMR experiments. The close correlation between the calculated (by molecular modelling) and observed ³J_{H–H} coupling constants of 3 (see the ESI†) suggests that the conformation adopted is similar to our modelled prediction.

The cell growth inhibitory activity of macrocyclic discodermolide/dictyostatin hybrid 3 was evaluated *in vitro* against three cancer cell lines: MDA-MB-231 (breast), A549 (non-small cell lung) and HT29 (colon) (Table 1). Notably, analogue 3 displayed significant antiproliferative activity against these human carcinoma cells, with a cytotoxicity around one-tenth that of discodermolide. This preliminary data is consistent with the conformation adopted by 3 closely resembling the X-ray structure of discodermolide, which itself has now been reported to correlate strongly with the NMR-derived conformation of discodermolide bound to tubulin.¹⁵



Scheme 2 Synthesis of enone 4 and aldehyde 5.



Scheme 4 Completion of the synthesis of hybrid analogue 3.

Table 1 Human cancer cell growth inhibitory properties of macrocyclic discodermolide analogue 3 relative to discodermolide (1)

	GI ₅₀ /μM ^a		
	MDA-MB-231 (breast)	A549 (non-small cell lung)	HT29 (colon)
1	0.029	0.020	0.015
3	0.208	0.399	0.170

^a 50% growth inhibitory concentration after 72 h of continuous incubation.

In conclusion, we have designed and synthesised the most active macrocyclic discodermolide/dictyostatin hybrid reported to date.^{7,16} The encouraging antiproliferative activity of analogue 3 can be attributed to its constrained (dictyostatin-like) macrocyclic structure, which bears a strong resemblance to the bioactive conformation of discodermolide.¹⁵ In ongoing work, this rational design approach is being extended to the synthesis of further novel hybrid analogues of discodermolide and dictyostatin.

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Notes and references

- (a) S. P. Gunasekera, M. Gunasekera, R. E. Longley and G. K. Schulte, *J. Org. Chem.*, 1990, **55**, 4912. CCDC reference VINTAN for the X-ray crystal structure of 1; (b) E. ter Haar, R. J. Kowalski, E. Hamel, C. M. Lin, R. E. Longley, S. P. Gunasekera, H. S. Rosenkranz and B. W. Day, *Biochemistry*, 1996, **35**, 243.
- R. J. Kowalski, P. Giannakakou, S. P. Gunasekera, R. E. Longley, B. W. Day and E. Hamel, *Mol. Pharmacol.*, 1997, **52**, 613.
- A. Mita, A. C. Lockhart, T.-L. Chen, K. Bochinski, J. Curtright, W. Cooper, L. Hammond, M. Rothenberg, E. Rowinsky and S. Sharma, *J. Clin. Oncol.*, 2004, **22**, 2025.
- G. S. Huang, L. L. Barcons, B. S. Freeze, A. B. Smith, III, G. L. Goldberg, S. B. Horwitz and H. M. McDaid, *Clin. Cancer Res.*, 2006, **12**, 298.
- (a) G. R. Pettit, Z. A. Cichacz, F. Gao, M. R. Boyd and J. M. Schmidt, *J. Chem. Soc., Chem. Commun.*, 1994, 1111; (b) R. A. Isbrucker, J. Cummins, S. A. Pomponi, R. E. Longley and A. E. Wright, *Biochem. Pharmacol.*, 2003, **66**, 75; (c) I. Paterson, R. Britton, O. Delgado and A. E. Wright, *Chem. Commun.*, 2004, 632; (d) I. Paterson, R. Britton, O. Delgado, A. Meyer and K. G. Poullennec, *Angew. Chem., Int. Ed.*, 2004, **43**, 4629.
- C. Madiraju, M. C. Edler, E. Hamel, B. S. Raccor, R. Balachandran, G. Zhu, K. A. Giuliano, A. Vogt, Y. Shin, J. Fournier, Y. Fukui, A. M. Brückner, D. P. Curran and B. W. Day, *Biochemistry*, 2005, **44**, 15053; R. M. Buey, I. Barasoain, E. Jackson, A. Meyer, P. Giannakakou, I. Paterson, S. Mooberry, J. M. Andreu and J. F. Diaz, *Chem. Biol.*, 2005, **12**, 1269.
- (a) Y. Shin, N. Choy, R. Balachandran, C. Madiraju, B. W. Day and D. P. Curran, *Org. Lett.*, 2002, **4**, 4443; (b) For an excellent review on natural product hybrids, see: L. F. Tietze, H. P. Bell and S. Chandrasekhar, *Angew. Chem., Int. Ed.*, 2003, **42**, 3996.
- (a) I. Paterson, G. J. Florence, K. Gerlach, J. P. Scott and N. Sereinig, *J. Am. Chem. Soc.*, 2001, **123**, 9535; (b) I. Paterson, O. Delgado, G. J. Florence, I. Lyothier, M. O'Brien, J. P. Scott and N. Sereinig, *J. Org. Chem.*, 2005, **70**, 150; (c) I. Paterson, O. Delgado, G. J. Florence, I. Lyothier, J. P. Scott and N. Sereinig, *Org. Lett.*, 2003, **5**, 35.
- M. S. Congreve, E. C. Davison, M. A. M. Fuhry, A. B. Holmes, A. N. Payne, R. A. Robinson and S. E. Ward, *Synlett*, 1993, 663.
- W. C. Still and C. Gennari, *Tetrahedron Lett.*, 1983, **24**, 4405.
- Attempts to remove the PMB ether using DDQ resulted in both PMP acetal formation and over-oxidation to the benzoate.
- D. A. Evans, K. T. Chapman and E. M. Carreira, *J. Am. Chem. Soc.*, 1988, **110**, 3560. NaBH(OAc)₃ gave 67 : 33 dr in favour of the 1,3-*syn*-diol while Me₂NBH(OAc)₃ gave little or no diastereoselectivity.
- E. J. Corey, R. K. Bakshi and S. Shibata, *J. Am. Chem. Soc.*, 1987, **109**, 5551.
- After chromatographic separation, the *syn*-diol was subsequently used to synthesise the C9 epimer of 3.
- V. M. Sanchez-Pedregal, K. Kubicek, J. Meiler, I. Lyothier, I. Paterson and T. Carlomagno, *Angew. Chem., Int. Ed.*, 2006, **45**, 7388.
- For recent reports of non-macrocyclic discodermolide analogues, see *inter alia*: I. Paterson and O. Delgado, *Tetrahedron Lett.*, 2003, **44**, 8877; A. B. Smith, III and M. Xian, *Org. Lett.*, 2005, **7**, 5229; A. B. Smith, III, B. S. Freeze, M. J. LaMarche, T. Hirose, I. Brouard, M. Xian, K. F. Sundermann, S. J. Shaw, M. A. Burlingame, S. B. Horwitz and D. Myles, *Org. Lett.*, 2005, **7**, 315; S. J. Shaw, K. F. Sundermann, M. A. Burlingame, D. C. Myles, B. S. Freeze, M. Xian, I. Brouard and A. B. Smith, III, *J. Am. Chem. Soc.*, 2005, **127**, 6532; J. M. Minguez, S. Y. Kim, K. A. Giuliano, R. Balachandran, C. Madiraju, B. W. Day and D. P. Curran, *Bioorg. Med. Chem.*, 2003, **11**, 3335; S. P. Gunasekera, R. E. Longley and R. A. Isbrucker, *J. Nat. Prod.*, 2002, **65**, 1830.